

SWINE HEALTH

Title: **Mechanism of PRRSV inhibition of interferon-mediated antiviral response – NPB #10-118**

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Scientific Abstract:

Porcine reproductive and respiratory syndrome virus (PRRSV) interferes with interferon (IFN)-activated antiviral response. In this study, different PRRSV strains were compared for their effects on IFN signal transduction pathway. One strain of genotype 1 PRRSV (LeLystad) and six strains of genotype 2 (VR-2385, Ingelvac PRRS MLV, VR-2332, NVSL, A2MC2 and MN-184) were used in this experiment. Compared to uninfected MARC-145 cells, all of the strains tested except A2MC2 and MN-184 had much lower protein level of IFN-stimulated gene 56 (ISG56) and signal transducer and activator of transcription 2 (STAT2) post-IFN treatment. Infection of the cells with VR-2385, MLV, VR-2332 and NVSL led to significant reduction of the transcript level of ISG15 and ISG56. In primary porcine alveolar macrophages (PAMs), all the strains except MLV and NVSL inhibited IFN-induced elevation of STAT2 protein. To determine which PRRSV protein(s) are responsible for the inhibition of the IFN signaling, individual proteins of VR-2385 were cloned for overexpression. IFN signaling luciferase reporter assay showed that in addition to nsp1 β , GP3, GP4 and N proteins inhibited the IFN-activated reporter expression. Nsp1 β protein was selected for further analysis and comparison between the different PRRSV strains for their effects on the IFN-activated JAK/STAT signaling. Nsp1 β from these strains were cloned into an expression vector and their effect on IFN signaling was compared. Overexpression of all the nsp1 β plasmids except MLV led to inhibition of the IFN reporter expression. This indicates that nsp1 β from all the strains except MLV has inhibitory effect on IFN signaling. Our findings delineate the different effects of these strains on IFN-activated signal transduction.

Type I IFNs induce the expression of IFN-stimulated genes by activating phosphorylation of both the STAT1 and STAT2, which form heterodimers, interact with IRF9 and translocate to the nucleus in heterotrimers (ISGF3). PRRSV nsp1 β was found to block the nuclear translocation of ISGF3 complex. We further discovered that nsp1 β induced degradation of karyopherin- α 1 (KPNA1, also known as importin- α 5). KPNA1 is known to mediate import of ISGF3 from cytoplasm to the nucleus. Overexpression of nsp1 β led to a reduction of KPNA1 but had little effect on its transcript level. Treatment of the nsp1 β -transfected cells with proteasome inhibitor MG132 restored KPNA1 protein level. Presence of nsp1 β led to elevation of KPNA1 ubiquitination and a shortening of its half-life. Analysis of nsp1 β deletion constructs showed that the N-terminal domain of nsp1 β was involved in the ubiquitin-proteasomal degradation of KPNA1. Infection of MARC-145 cells by PRRSV strain VR-2332 and VR-2385 led to a reduction of KPNA1, while a low virulent strain MLV had little effect. These results indicate that nsp1 β blocks ISGF3 nuclear translocation by way of stimulating KPNA1 degradation.

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